Diff Quik® Staining Protocol

Materials:

Fixative: Methanol

Stain 1: Eosinophilic (red) stain

Stain 2: Basophilic (blue) stain

Other: Frosted slides, coverslips, pencils, forceps, gloves, immersion oil, lens tissue, plastic pipettes, cotton-tips, lens cleaning fluid, +/- ionised water, absorbent material or paper towel, slide cases, storage container such as a lunch box for slides, coverslips, pencils etc.

Protocol:

1. Ensure sample is labelled in pencil with the patient's name, sample site, and date. Confirm that the sample is on the same side as the writing.

2. Fix. Most samples will need to stand in the fixative for at least 30 seconds. Tap gently on absorbent paper towel to remove access fixative.

3. Red stain: Dip slide into red stain 5-10 times, each dip lasting one second. Tap gently on absorbent paper towel to remove access stain.

4. Blue stain: Dip slide into blue stain 5-10 times, each dip lasting one second. Tap gently on absorbent paper towel to remove access stain.

5. Rinse: Gently run water over the slide to rinse off the access stain.

6. Prop slide upright either in a slide rack or leaning against something secure at the Diff Quik® station. You may gently fan the slide to speed up air drying. Avoid using a hair dryer on fine needle aspirates or blood films.

7. Prior to viewing, place a small drop of immersion oil onto the stained area of the slide and then add a cover slip. The slide is now ready for viewing.

Stain maintenance:

Dirty stains to be changed at least once weekly, more often if required. Last changed on: _____

Clean stains to be changed once weekly. Last changed on: _____